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(54) Title: HUMANIZATION OF AN ANTI-CARCINOEMBRYONIC ANTIGEN ANTI-IDIOTYPE ANTIBODY AND USE AS A  
TUMOR VACCINE AND FOR TARGETING APPLICATIONS

(57) Abstract

A humanized form of an anti-idiotype antibody to CEA, e.g., hW12, has conserved immunoreactivity. The clinical benefits of anti-CEA antibodies are maximized by using the humanized anti-idiotype as a clearing agent for anti-CEA antibodies or antibody fragments. The humanized anti-idiotype also can be used as an immunogenic vaccine.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide a chimeric anti-idiotypic Ab with anti carcinoembryonic-antibody properties, a humanized anti-idiotypic Ab with anti carcinoembryonic-antibody properties as an immunological reagent useful in clearing an organism of anti-CEA antibody initially used as a cancer treatment, diagnostic, or vaccine, where the anti-idiotypic Ab has the immunogenic properties of a human MAb in a human patient, and to provide an anti-idiotypic Ab with anti carcinoembryonic-antibody properties which can serve as a detection agent or vaccine. It is another object of the present invention to provide DNA constructs encoding such antibodies.

To achieve these objectives, in one aspect of the invention, a chimeric anti-idiotypic antibody or fragment thereof which specifically binds to the idiotype region of an anti-CEA monoclonal antibody is provided, comprising the rWI2 light chain and heavy chain variable regions, or silent mutations thereof. In a preferred embodiment, the heavy chain variable region comprises the ratWI2VK sequence shown in Figure 1 and the light chain variable region comprises the RatWI2VK sequence shown in Figure 2.

In another aspect of the invention, a humanized anti-idiotypic antibody or fragment thereof which specifically binds the idiotype region of an anti-CEA monoclonal antibody is provided, comprising rWI2 CDR regions and humanized FR regions. In a preferred embodiment, the heavy chain variable region comprises the KOLWI2VH-1 or the KOLWI2VH-2 sequence shown in Figure 1 and the light chain variable region comprises the REIWI2VK or the REIWI2VKRS sequence shown in Figure 2.

comprising a gene for a humanized WI2 light chain are provided.

5 In another aspect of the invention, a host is provided, comprising an isolated first vector comprising a gene for the chimeric WI2 heavy chain and an isolated second expression vector comprising a gene for the chimeric WI2 light chain, or an isolated first expression vector comprising a gene for a humanized WI2 heavy chain and an isolated second expression vector comprising a gene for a humanized WI2 light chain.

15 In another aspect of the invention, a method of stimulating an immune response in a patient against cancers expressing carcinoembryonic antigen is provided, which comprises administering to a patient an effective amount of a vaccine comprising a humanized anti-idiotypic antibody or fragment which specifically binds the idiotype region of an anti-CEA monoclonal antibody, conjugated to a soluble immunogenic carrier protein, optionally in combination with a pharmaceutically acceptable vaccine adjuvant.

25 In another aspect of the invention, a method of diagnosis or treatment of a patient is provided, wherein an antibody or antibody fragment that specifically binds CEA is used as a targeting, pre-targeting or therapy agent, either as such or as a component of a conjugate, the improvement to the method consisting of an anti-idiotypic antibody used to clear non-targeted antibody or antibody fragment.

30 In another aspect of the invention, a method of detecting the presence of an antibody or fragment that specifically binds CEA, in a fluid biological sample, comprising contacting the sample with rWI2, or a chimeric anti-idiotypic antibody or antibody fragment which specifically binds the idiotype region of an anti-CEA

monoclonal antibody, or a humanized anti-idiotypic antibody or antibody fragment which specifically binds the idiotype region of an anti-CEA monoclonal antibody, and detecting binding of said anti-idiotypic antibody or antibody fragment to an antibody idiotype or antibody idiotype fragment in said sample.

#### DESCRIPTION OF THE FIGURES

Figure 1 shows the design of the hVH. The rvH is shown aligned with KOL, and with the designed KOLW2VH-1 and 2 sequences. Dashes indicate that the sequence matches at that position the rWI2VK sequence. Narrow boxes indicate positions where the rat aa was retained in the humanized FR sequences. Note that the KOLWI2VK-1 sequence contains an extra rat aa, when compared with KOLWI2VK-2, at position 5. CDRs are indicated by wider boxes and are labeled above the respective box as CDR1 to CDR3.

Figure 2 shows the design of hVK. The RATWI2VK sequence is shown aligned with REI, and with the designed REIWI2VK and REIWI2VKRS. Dashes indicate that the sequence matches at that position the RATWI2VK sequence. Narrow boxes indicate positions where the rat aa was retained in the humanized FR sequences. As indicated, four rat aa residues were retained in the FR regions. CDRs are indicated by wider boxes and are labeled above the respective box as CDR1-3.

Figure 3 shows the polynucleotide sequence for the hWI2 heavy chain variable region. The PCR primers employed and the synthesized oligo K and oligo L described in the text are indicated. A single aa letter code is given below the polynucleotide sequence to represent the translation product. The CDRs are underlined on the protein sequence.

Figure 4 shows the polynucleotide sequence for the hWI2 light chain variable region. The PCR primers employed and the synthesized oligo M' and oligo N described in the text are indicated. A single aa letter code is given below the polynucleotide sequence for the translation product. The CDRs are underlined on the protein sequence.

Figure 5 shows a comparison between hWI2(RS), hWI2, cWI2, and rWI2 in competitive binding assays. Binding of each of these antibodies is shown as a percentage of inhibition of the binding by a peroxidase conjugated MN-14 antibody.

Figure 6 shows the nucleic acid sequence for the variable region of rWI2 light chain. The protein translation product is indicated below the nucleic acid sequence, using one letter aa code. The aa residues representing CDRs 1-3 are underlined and labeled.

Figure 7 shows the nucleic acid sequence for the variable region of rWI2 heavy chain. The protein translation product is indicated below the nucleic acid sequence, using one letter aa code. The aa residues representing CDRs 1-3 are underlined and labeled.

#### ILLUSTRATIVE GLOSSARY

The following terms or abbreviations are used in the present application. The meanings set out in this glossary are for illustrative purposes only. The full meaning of the terms will be apparent to those of skill in the art.

"CDR" is used as an abbreviation for Complementarity Determining Region. These are the regions within the variable regions of an antibody that are primarily, but

An isolated DNA molecule is a fragment of DNA that is not integrated in the genomic DNA of an organism. For example, a cloned T cell receptor gene is a DNA fragment that has been separated from the genomic DNA of a mammalian cell. Another example of an isolated DNA molecule is a chemically-synthesized DNA molecule that is not integrated in the genomic DNA of an organism.

Complementary DNA (cDNA) is a single-stranded DNA molecule that is formed from an mRNA template by the enzyme reverse transcriptase. Typically, a primer complementary to portions of mRNA is employed for the initiation of reverse transcription. Those skilled in the art also use the term "cDNA" to refer to a double-stranded DNA molecule consisting of such a single-stranded DNA molecule and its complementary DNA strand.

The term expression refers to the biosynthesis of a gene product. For example, in the case of a structural gene, expression involves transcription of the structural gene into mRNA and the translation of mRNA into one or more polypeptides.

A cloning vector is a DNA molecule, such as a plasmid, cosmid, or bacteriophage, that has the capability of replicating autonomously in a host cell. Cloning vectors typically contain one or a small number of restriction endonuclease recognition sites at which foreign DNA sequences can be inserted in a determinable fashion without loss of an essential biological function of the vector, as well as a marker gene that is suitable for use in the identification and selection of cells transformed with the cloning vector. Marker genes typically include genes that provide tetracycline resistance or ampicillin resistance.

An expression vector is a DNA molecule comprising a gene that is expressed in a host cell. Typically, gene

An antibody fragment is a portion of an antibody such as F(ab')<sub>2</sub>, F(ab)<sub>2</sub>, Fab', Fab, and the like. Regardless of structure, an antibody fragment binds with the same antigen that is recognized by the intact antibody. For example, an anti-CEA Mab (Ab1) fragment binds with CEA, while an Ab2 fragment binds with the variable region of the Ab1 and mimics an epitope of CEA.

The term "antibody fragment" also includes any synthetic or genetically engineered protein that acts like an antibody by binding to a specific antigen to form a complex. For example, antibody fragments include isolated fragments consisting of the light chain variable region, "Fv" fragments consisting of the variable regions of the heavy and light chains, recombinant single chain polypeptide molecules in which light and heavy variable regions are connected by a peptide linker ("sFv proteins"), and minimal recognition units consisting of the amino acid residues that mimic the hypervariable region.

As used herein, the term antibody component or the term antibody may include both an entire antibody and an antibody fragment.

Vaccine as used in the present context is an antibody or antibody fragment which is engineered to more effectively stimulate an immune response. Typically, this is achieved by conjugation of the antibody component to a soluble immunogenic carrier protein.

#### DETAILED DESCRIPTION OF THE INVENTION

An overview. Providing chimeric and humanized WI2 anti-idi typ antibodies.

Rat WI2 (rWI2) is an anti-idiotypic monoclonal antibody which binds the CDR of MN-14. See Losman et

al., (1994) *supra*. It was previously proposed that PCR primer pairs may be designed which might isolate the variable regions of rat antibodies from a RNA substrate. See Kutemeier et al., *Hybridoma*, 11: 23-32 (1992).  
5 Similar approaches have been employed to isolate the variable regions of mouse antibodies. See Orlandi et al., *Proc. Nat'l Acad. Sci. USA* 86: 3833 (1989). In the present invention, a combination of PCR primers as described by Kutemeier et. al., and by Orlandi et al.,  
10 *supra*, are required to isolate the rWI2 variable region. These primers could be used to isolate other rat variable regions.

cDNA for the variable regions of both the light and heavy chains of rWI2 are isolated and the sequence from  
15 a number of clones is confirmed. The rat variable regions are cloned into plasmid vectors, such that the rat variable K and H regions are attached to respective human IgG constant regions. The plasmids are transfected into SP2/O cells, clones comprising both plasmids are  
20 selected, and chimeric WI2 (cWI2) antibody is expressed.

To humanize the variable regions, FR rat regions are replaced by the human IgG counterparts. The rat CDR regions are preserved, in order to maintain the specificity of the antibody. However, to enhance  
25 stability and maintain specificity and binding affinity of the antibody, it is best to employ a human sequence which is most similar to the newly identified rat FR sequences. Rat residues that are close to the CDRs, or known from prior experience to be important for  
30 interactions with the CDRs, were retained in both humanized sequences.

For both the heavy and light chain, long oligonucleotides corresponding to the variable region are engineered to reflect the humanized design. The  
35 oligonucleotides replace the rat variable region in



plasmid constructs designated to produce full length human WI2 (hWI2). The effectiveness of cWI2 or hWI2 are compared with that of rWI2 in assays where they compete with CEA for binding to a peroxidase conjugated MN-14.

5        In order to express the cWI2 and the hWI2s, the rat and the humanized variable regions are inserted into one or separate plasmid vectors in such a manner as to allow coexpression of full length light and heavy antibody chains. The vectors can be transfected into SP2/0 cells.  
10       Selection of transfected cells, and eventual amplification of the clones, are based on the expression of the *dihydrofolate reductase* (DHFR) gene which is located on the same plasmid vector. The functionality of various constructs is determined by binding assays. For  
15       example, final hWI2 constructs were compared with both rWI2 and cWI2 in their ability to bind MN-14 -- see Figure 5.

#### Production of Monoclonal Ab1 and Ab2 Antibodies

20       Rodent monoclonal antibodies to specific antigens may be obtained by methods known to those skilled in the art. See, for example, Kohler and Milstein, *Nature* 256: 495 (1975), and Coligan et al. (eds.), *CURRENT PROTOCOLS IN IMMUNOLOGY*, VOL. 1, pages 2.5.1-2.6.7 (John Wiley & Sons 1991) [hereinafter "Coligan"]. Briefly, monoclonal  
25       antibodies can be obtained by injecting rats with a composition comprising an antigen, verifying the presence of antibody production by removing a serum sample, removing the spleen to obtain B-lymphocytes, fusing the B-lymphocytes with myeloma cells to produce hybridomas,  
30       cloning the hybridomas, selecting positive clones which produce antibodies to the antigen, culturing the clones that produce antibodies to the antigen, and isolating the antibodies from the hybridoma cultures. These techniques are applicable for production of a monoclonal antibody  
35       from other mammals injected with an antigen.

A wide variety of monoclonal antibodies against tumor associated antigens or infectious agents have been developed. See, for example, Goldenberg et al., international application publication No. WO 91/11465 (1991), and Goldenberg, international application publication No. WO 94/04702 (1994), each of which is incorporated herein by reference in its entirety.

Polyclonal Ab2 can be prepared by immunizing animals with Ab1 or fragments, using standard techniques. See, for example, Green et al., "Production of Polyclonal Antisera," in METHODS IN MOLECULAR BIOLOGY: IMMUNOCHEMICAL PROTOCOLS, Manson (ed.), pages 1-12 (Humana Press 1992). Also, see Coligan at pages 2.4.1-2.4.7. Alternatively, monoclonal Ab2 can be prepared using Ab1 or fragments as immunogen with the techniques described above.

MABs can be isolated and purified from hybridoma cultures by a variety of well-established techniques. Such isolation techniques include affinity chromatography with Protein-A Sepharose, size-exclusion chromatography, and ion-exchange chromatography. See, for example, Coligan at pages 2.7.1-2.7.12 and pages 2.9.1-2.9.3. Also, see Baines et al., "Purification of Immunoglobulin G (IgG)," in METHODS IN MOLECULAR BIOLOGY, VOL. 10, pages 79-104 (The Humana Press, Inc. 1992).

#### The Design of a Humanized Antibody

One antibody of the present invention is a "humanized" monoclonal antibody. That is, rat complementarity determining regions are transferred from heavy and light variable chains of the rat immunoglobulin into a variable region designed to contain a number of aa residues found within the FR region in human IgG. Similar conversion of mouse/human chimeric antibodies to a humanized antibody has been described before. General techniques for cloning murine immunoglobulin variable

domains are described, for example, by the publication of Orlandi et al., *Proc. Nat'l Acad. Sci. USA* 86: 3833 (1989), which is incorporated by reference in its entirety. Techniques for producing humanized MAbs are described, for example, by Jones et al., *Nature* 321: 522 (1986), Riechmann et al., *Nature* 332: 323 (1988), Verhoeyen et al., *Science* 239: 1534 (1988), Carter et al., *Proc. Nat'l Acad. Sci. USA* 89: 4285 (1992), Sandhu, *Crit. Rev. Biotech.* 12: 437 (1992), and Singer et al., *J. Immun.* 150: 2844 (1993), each of which is hereby incorporated by reference.

#### Construction of cWI2 and hWI2.

The engineering of chimeric and humanized WI2 employs standard molecular biology and cellular biology approaches, for example PCR reactions, sequencing of DNA, synthesis of long oligonucleotides, cloning, site-directed mutagenesis, transfection of vectors into cells, expression of chimeric and humanized antibodies, their purification and so on. The techniques employed are standard techniques, well known in the art and well established. A person skilled in the art would have no difficulty carrying out those techniques. All necessary materials are readily available. Reference manuals describing these techniques are widely available. For example, see Sambrook et al., *Molecular Cloning: A Laboratory Manual*, second ed., Cold Spring Harbor Press, (1989); Co et al., *J. Immunol.*, 148: 1149 (1992); Ausubel et al., eds., *CURRENT PROTOCOLS IN MOLECULAR BIOLOGY*, and updates, John Wiley & Sons, NY, (1987); "Protocols - Current Methods and Applications" White, ed., *Methods in Mol. Biol.* (15), Humana Press, Totawa, New Jersey (1993); Uhlmann, *Gene* 71: 29 (1998); Wosnick et al., *Gene* 60: 115 (1988); and Huse, in *ANTIBODY ENGINEERING: A PRACTICAL GUIDE*, Boerrbaeck, ed., W.H. Freeman & Co., 103-120 (1992).

The PCR primers used in the initial cloning of the rat variable region are designed over regions of the sequence that are expected to be relatively conserved between human antibodies. Furthermore, an amino acid (aa) can generally be represented by more than one codon, although the codons often differ only in the third position. Such codons are sometimes referred to as "degenerate." To design the PCR primer one attempts to choose a protein sequence over which degenerate codons are limited in number. The approach is further limited by the need to design over a conserved protein sequence, as discussed above. Therefore, often, the "primer" as designed actually consists of a mix of numerous molecules that differ in sequence at specific positions. Any one primer molecule, which would provide a perfect match for the specific mRNA substrate to be amplified is present as a minuscule fraction of the overall primer mix. To enhance likelihood of success, a few primer pairs, i.e. covering different regions of the mRNA are generally tried.

One skilled in the art recognizes that alternative techniques are readily available. For example, initial changes to the cloned rat variable region could be introduced by any of a number of site specific mutagenesis protocols. Furthermore, it should be realized that additional aa changes in the variable and especially the FR regions may be possible, with the expectation that there will be some changes which would be silent in nature. Silent changes are those replacements, deletion or addition of one or a small number of aa that would not significantly affect the binding affinity or specificity of the antibody or antibody fragments thereof. Most obvious among such silent changes would be the replacement of one aa by an aa of a similar size and chemical properties. Such changes are well known in the art and are generally referred to as a "conservative" aa substitution. For

example, a leucine when replaced by an isoleucine would generally not be expected to affect the structure of a protein. All such conserved aa changes, as well as silent changes which do not significantly affect the binding affinity or specificity of the anti-carcinoembryonic antibody, are within the scope of the invention.

#### **Expression of the Engineered cWI2 or hWI2**

Genes encoding the antibodies of the invention are introduced via expression vectors into a host cell, for expression. In a preferred embodiment, the genes for both the light and heavy genes are introduced in a single expression vector, which is introduced in a host cell. The expression vectors generally contain drug markers for selection of the transformed cell. A drug marker can furthermore be used to amplify the copy number of nearby genes, resulting in a clone overexpressing the antibody. For example, a vector expressing the light and heavy chains of cWI2 or hWI2 were introduced into SP2/0 cells on vectors containing the DHFR gene. The original clones were amplified after selection by growth on methotrexate (MTX).

It should be understood that alternative ways to coexpress the light and heavy chain genes are feasible. A skilled artisan could consider other selection regimens, introduction of both the light and heavy chain genes on one plasmid or cotransformation with separate vectors encoding the light and heavy genes, and transfection of other cell lines. Furthermore, expression of the antibody in yet other systems is possible. For example, expression could occur in yeast. Alternatively, baculoviruses can be engineered with the light and heavy genes and expressed in cultured cells, or used to infect an insect.

The antibodies require purification from their expression system and media by methods that are generally similar to methods described above for purification of MAbs from hybridomas.

5           **Uses for the Antibodies of the Invention**

Humanized monoclonal antibodies in accordance with the invention are suitable for use in therapeutic methods. For example, MN-14 has been proposed as a therapeutic agent or as a vaccine to stimulate Ab2. The  
10 dosage of MN-14 conjugated to a drug can be enhanced if removal or clearing of unbound MN-14 is made possible by its binding, and aggregate formation, with hWI2. Similarly, MN-14 can be used as a vaccine, optionally by alternative applications with an anti-idiotypic antibody  
15 vaccine. Delivery of the Ab2 would be counterproductive in the presence of excess, free-floating MN-14 vaccine. Here, hWI2, for example, would clear the MN-14 vaccine, to allow efficient administration of the Ab2 vaccine.

In addition, hWI2 itself can be a vaccine.  
20 Generally, the antibodies and fragments of the present invention can be used as vaccines by conjugating the antibodies or fragments to a soluble immunogenic carrier protein. Suitable carrier proteins include keyhole  
25 lympet hemocyanin, which is the preferred carrier protein. The antibodies and fragments can be conjugated to the carrier protein using standard methods. See, for example, Hancock et al, "Synthesis of Peptides for Use as Immunogen," in METHODS IN MOLECULAR BIOLOGY:  
IMMUNOCHEMICAL PROTOCOLS, Manson (ed.), pages 23-32  
30 (Humana Press 1992).

A preferred vaccine composition comprises an antibody conjugate or fragment conjugate, and an adjuvant. Examples of suitable adjuvant include aluminum hydroxide and lipid. Methods of formulating vaccine compositions  
35 are well-known to those of ordinary skill in the art.

See, for example, Rola, "Immunizing Agents and Diagnostic Skin Antigens," in REMINGTON'S PHARMACEUTICAL SCIENCES, 18th Edition, Gennaro (ed.), pages 1389-1404 (Mack Publishing Company 1990).

5 Additional pharmaceutical methods may be employed to control the duration of action of a vaccine in a therapeutic application. Controlled release preparations can be prepared through the use of polymers to complex or adsorb the antibodies or fragments. For example, bio-  
10 compatible polymers include matrices of poly(ethylene-co-vinyl acetate) and matrices of a polyanhydride copolymer of a stearic acid dimer and sebacic acid. Sherwood et al., *Bio/Technology* 10: 1446 (1992). The rate of release of an antibody or antibody fragment from such a matrix  
15 depends upon the molecular weight of the antibody or fragment, the amount of antibody or fragment within the matrix, and the size of dispersed particles. Saltzman et al., *Biophys. J.* 55: 163 (1989); Sherwood et al., *supra*. Other solid dosage forms are described in Ansel et al.,  
20 PHARMACEUTICAL DOSAGE FORMS AND DRUG DELIVERY SYSTEMS, 5th Edition (Lea & Febiger 1990), and Gennaro (ed.), REMINGTON'S PHARMACEUTICAL SCIENCES, 18th Edition (Mack Publishing Company 1990).

25 The anti-idiotypic Ab of the invention can be used for the detection of an anti-carcinoembryonic antibody. For example, rWI2, cWI2 and hWI2 can be used *in vitro* to test the blood sample of a patient for the presence of anti-carcinoembryonic antibody. Detection of MN-14 levels, when MN-14 is used as a therapeutic agent or as a vaccine  
30 would be important. Presence of MN-14 can be detected by rWI2, cWI2, r hWI2. Similarly, hWI2 can be used in a patient to determine the presence of natural anti-CEA Abs. To be useful in the detection of an anti-CEA Ab, the anti-idiotypic Abs can be conjugated to a label.  
35 Suitable labels include, e.g., a radiolabel, an enzyme,

or a fluorescent label. Such labeling agents and methods of conjugation are well known to one skilled in the art.

The antibody preparations of the present invention can be formulated according to known methods to prepare pharmaceutically useful compositions, whereby antibodies or antibody fragments are combined in a mixture with a pharmaceutically acceptable carrier. A composition is said to be a "pharmaceutically acceptable carrier" if its administration can be tolerated by a recipient mammal. Sterile phosphate-buffered saline is one example of a pharmaceutically acceptable carrier. Other suitable carriers are well-known to those in the art. See, for example, Ansel et al., PHARMACEUTICAL DOSAGE FORMS AND DRUG DELIVERY SYSTEMS, 5th Edition (Lea & Febiger 1990), and Gennaro (ed.), REMINGTON'S PHARMACEUTICAL SCIENCES, 18th Edition (Mack Publishing Company 1990).

The antibodies or fragments may be administered to a mammal intravenously or subcutaneously. Moreover, the administration may be by continuous infusion or by single or multiple boluses. Preferably, an antibody vaccine is administered subcutaneously, while an antibody preparation that is not a vaccine is administered intravenously. In general, the dosage of administered antibodies or fragments for humans will vary depending upon such factors as the patient's age, weight, height, sex, general medical condition and previous medical history. Typically, it is desirable to provide the recipient with a dosage of antibodies or fragments which is in the range of from about 1 pg/kg to 10 mg/kg (amount of agent/body weight of patient), although a lower or higher dosage also may be administered as circumstances dictate.

For purposes of therapy, antibodies or fragments are administered to a mammal in a therapeutically effective amount. An antibody preparation is said to be



(RVK-3BACK) 5'-CCA GAT GTG AGC TCG TGA TGA CCC AGA CTC  
CA-3',  
(RVK-4BACK) 5'-CCA GAT GTG AGC TCG TCA TGA CCC AGT CTC  
CA-3',  
5 (RVK-5BACK) 5'-CCA GTT CCG AGC TCG TGA TGA CAC AGT CTC  
CA-3', and  
(RVK-1FOR) 5'-GTT AGA TCT CCA GCT TGG TCC C-3'.

#### The cloning of VH sequence

First-strand cDNA was prepared from total RNA  
10 isolated from WI2 hybridoma using random hexamers as the  
annealing primers. RCH1 (which anneals to the rat IgG1  
CH1 domain) and RVH-1BACK (which anneals to rat 5' VH  
region) were tried as a primer pair to PCR-amplify and  
isolate the VH sequence from the first-strand cDNA  
15 template, using standard protocols. However, no PCR  
product was obtained. Unexpectedly, the RCH1 primer, in  
conjunction with the Orlandi primer VH1BACK (5'-AGG TSM  
ARC TGC AGS AGT CWG G-3') produced a PCR product of the  
expected size. The PCR-amplified VH sequence was  
20 digested with the restriction enzymes PstI/BstEII and  
subcloned into the corresponding sites of the heavy chain  
staging vector, VHpBS. See Leung et al., *Hybridoma* 13:  
469 (1994). Six individual clones were sequenced and  
confirmed to be identical to each other. RVH-1FOR which  
25 anneals to rat 3' VH region and RVH-1BACK also produced  
a PCR product of the expected size, which was not further  
analyzed. Figure 7 shows the sequence of the heavy chain  
variable region.

#### The cloning of VK sequence

30 First-strand cDNA was prepared from total RNA  
isolated from WI2 hybridoma using random hexamers as the  
annealing primers. The RK1 (which anneals to rat CK  
domain) in conjunction with RVK-3BACK (which anneal to  
the 5' VK region), was used to PCR-amplify and isolate  
35 the VK region of rWI2, employing standard PCR protocols.  
The expected 320 base pairs (bp) product was obtained.  
The PCR product was further analyzed. Other primer pairs

were not tested. Figure 6 shows the sequence of the light chain variable region.

Restriction analysis of the 320 bp VK PCR product revealed the presence of an additional PvuII site which would interfere with subsequent cloning into the PvuII/BcII sites of the staging vector, VKpBR. *Id.* Because the PCR product lacked the appropriate ends for the staging vector, it was directly subcloned into the TA Cloning Vector (Invitrogen), which allowed for direct insertions of PCR DNA. Six individual clones were sequenced and all six were shown to be identical. Analysis of the deduced protein sequence indicated a functional VK domain. To subclone the sequence into a staging vector, a new primer was designed. In this primer, a PvuII compatible end, rather than the whole restriction recognition site, was introduced so that the PCR product would have on its 5' end a compatible site for ligation into the corresponding PvuII site within the VKpBR staging vector. The PCR product was digested with BglII and subcloned into the PvuII/BcII cloning sites of the VKpBR staging vector. *Id.* The ligation was transformed in *E. coli*, and plated on selective plates. Miniprep DNA was prepared and analyzed. DNA from positive clones were sequenced to make sure that the joining region, at the PvuII site, was as expected. The PCR-amplified VH and VK sequence for WI2 were excised from their respective staging vector by HindIII/BamHI restriction digestion and the isolated fragments were subcloned into their respective heavy and light chain expression vectors, pGlg and pKh. *Id.* Chimeric WI2 (cWI2) was purified from clones cotransfected with the heavy and light chain expression vectors and the immunoreactivity of cWI2 was compared to that of rat WI2. Results indicated that both antibodies inhibited the binding of MN-14 onto CEA to a similar extent, confirming the functionality of the chimeric antibody, and the authenticity of the VH and VK sequences.

## Design of the Humanized WI2 Sequence

### VH and VK design

By comparing the sequence homology of a number of human IgG to the rWI2 frame work region, the human KOL framework was chosen for grafting the heavy chain CDRs. Two versions of the humanized heavy chain designated as KOLWI2VH-1 and KOLWI2VH-2 were designed. See Figure 1. However, only the KOLWI2VH-1 was synthesized and tested. KOLWI2VH-1 differs from KOLWI2VH-2 only by one amino acid at position 5(Q instead of V). Rat residues that are close to the CDRs, or known from prior experience to be important for interactions with the CDRs, were retained in both humanized sequences. A total of 5 rat residues were retained in the FR region of the humanized WI2 heavy chain sequence KOLWI2VH-1. The KOLWI2VH-2 sequence would have retained four rat aa residues.

The REI framework was chosen for grafting the light chain CDRs. Four rat residues were retained in the FR region of the designed light chain. The sequence of the designed light chain, REIWI2VK is shown in Figure 2.

### Gene synthesis for humanized WI2 VH and VK domain

Using computer analysis, the nucleotide sequences encoding the humanized VH and VK domain of WI2 were assembled as follows.

#### 25 hWI2 VH

The long oligo K (135-mer) was PCR-amplified using the flanking primers oligo 21 (45-mer) and oligo 22 (54-mer). The PCR product for oligo K encodes for the N-terminal half of the VH domain. Similarly, the long oligo L (133-mer) was PCR-amplified using the flanking primers oligo 23 (48-mer) and oligo 24 (45-mer). The PCR product for oligo L encodes for the C-terminal half of the VH domain. The sequences of the different oligos used for the PCR-synthesis of the humanized WI2 VH domain are summarized in Figure 3.

The PCR product for oligo K was digested with PstI/AlwNI, whereas the PCR product for oligo L was digested with AlwNI/BstEII. The K and L fragments were gel-purified and ligated to the PstI/BstEII site of the staging, vector VHpBR. Minipreps were analyzed by restriction analysis as well as by sequencing reaction. MP33#4(1.6.95) was confirmed to have the correct sequence for the hWI2 VH, and was used for subsequent cloning and subcloning. The BamHI/HindIII fragment containing the hWI2 VH domain was gel-purified from the DNA after restriction digestion, and was cloned into the corresponding sites of the heavy chain expression vector, pGlg. Leung et al. *Hybridoma* 13:469 (1994).

#### hWI2 VK

The long oligo M' (129-mer) was PCR-amplified using the flanking primers oligo 25 containing a PvuII site and oligo 26. The PCR product for oligo M' encodes for the N-terminal half of the VK domain. The long oligo N (127-mer) was PCR-amplified by the flanking primers oligo 27 (44-mer) and oligo 28 (39-mer). The PCR product for oligo N encodes for the C-terminal half of the VK domain. The sequences of the oligonucleotides used for the PCR-amplification of the humanized WI2 VK domain are summarized in Figure 4.

The PCR product for oligo M' was digested with PstI and PvuII, whereas the PCR product for oligo N was digested with PstI/BglII. The M and N fragments were gel-purified and ligated to the PvuII/BcII site of the staging vector VKpBR. By analysis of miniprep DNA three clones were identified to be the likely positives for hWI2VKpBR. Hogness stocks for these three minipreps were prepared. The sequence was confirmed from both directions, crossing the PvuII and BglII/BcII junctions. Plasmid DNA was prepared. The BamHI/HindIII fragment containing the hWI2 VK domain was gel-purified from the DNA after restriction digestion, and was cloned into the

corresponding sites of the light chain expression vector, pKh. Id. Plasmid DNA for both the heavy and light chain vectors expressing hWI2 were prepared and cotransfected into SP2/O myeloma cells by electroporation.

5       A total of eight antibody producing colonies were identified using ELISA assays. The three highest producers were maintained and cell aliquots were frozen for storage. The highest producer was expanded and upscaled for subsequent antibody purification and  
10       testing.

**Examining the immunoreactivity of hWI2 heavy chain in a mix-and-match experiment**

The mix-and-match antibody containing the humanized heavy and chimeric light chains of WI2 was purified.  
15       Various concentrations of the mix-and-match antibody were used to compete with ELISA-plate coated CEA for binding to the peroxidase-conjugated MN-14 antibody. Chimeric Ab was produced and expressed and shown to have immunogenic reactivity comparable to that of the rat WI2. The  
20       immunoreactivity of the mix-and-match WI2 was compared to that of cWI2, which has been confirmed to be identical to that of rat WI2. The humanized heavy chain for WI2 maintained the original immunoreactivity of rat WI2.

**WI2**

25       The original sequences for humanized WI2 heavy and light chains were designed from sequence homology comparison without the aid of computer modeling (Figure 2). In a competitive binding assay, the first version of humanized WI2 (hWI2) was shown to be slightly less  
30       effective in blocking the binding of peroxidase conjugated MN-14 to CEA, as compared to that of rWI2 and cWI2 (Figure 5). In the mix-and-match experiment (rVK + hVH) described above, it was demonstrated that the design of the humanized heavy chain was good. Therefore, a  
35       redesign of the humanized VK sequence was undertaken in

an attempt to restore the full immunoreactivity of hWI2. Studies on the crystal structure for REI protein suggested that the rat residues Arg (66) and Ser (69) might be interacting with the light chain CDRs. These  
5 two rat residues, arg and ser were reintroduced into the VK FR3 of hWI2 by site-directed mutagenesis. The newly designed light chain, REIWI2VKRS is shown in figure 2. Humanized WI2 containing these two mutations was designated as hWI2RS. All mutations and vector  
10 constructions were confirmed by sequencing as well as by extensive restriction analysis. Expression vectors for this new version of humanized light chain (hWl2RSpKh) were used for transfection experiments by electroporation.

15 For unknown reasons, it had been difficult to get antibody producing clones transfected with the hWl2RS expression vector. Clones 131A9 and 132F7 were identified as positive clones producing about 0.4 and 0.2 mg/liter of hWl2RS antibodies, respectively. One liter  
20 cultures of clone 131A9 and 132F7 were grown in roller bottles and the antibody was purified by a standard protein purification method. Approximately 0.4 and 0.2 mg of hWl2RS was obtained from clone 131A9 and 132F7, respectively. The immunoreactivities of 131A9 and 132F7  
25 hWl2RS, as measured by their abilities to block MN-14 binding on CEA, were shown to be similar.

Example 2**Comparing the immunoreactivities of different versions  
of WI2**

In one experiment, the immunoreactivities of hWI2 and  
5 hWI2RS, as measured by their abilities to block MN-14  
binding on CEA, were compared to that of rWI2 and cWI2,  
respectively (Figure 5). Both hWI2 and hWI2RS appeared  
to exhibit a slightly reduced immunoreactivities than  
that of rWI2 and cWI2. The introduction of the RS  
10 mutation into the FR3 region of hWI2 did not seem to  
improve immunoreactivity as predicted. Nevertheless,  
hWI2 was nearly as immunoreactive as rWI2 or cWI2.

Both hWI2 and hWI2RS exhibited very similar, if not  
identical, immunoreactivity. Since hWI2RS contains two  
15 extra rat residues in the light chain FR3 region, and  
would be potentially more immunogenic the first humanized  
version of WI2, hWI2, was chosen to be the antibody that  
was further scaled up for production.

Example 3**20 Construction of an amplifiable expression vector for  
hWI2**

The VH and VK sequence for hWI2 were excised from the  
respective staging vectors and ligated to a single  
amplifiable vector, pdHL2. The pdHL2 vector contains  
25 sequences for the human CK, IgG1, and an amplifiable DHFR  
gene, each controlled by separate promoters. See Leung  
et al., *Tumor Targeting* 2:184 (#95) (1996) and Losman et  
al., *Tumor Targeting* 2:183 (#93) (1996). The resultant  
vector expressing the hWI2 was designated hWI2pdHL2. The  
30 plasmid DNA for hWI2pdHL2 was linearized and transfected  
by electroporation into SP2/O cells. Selection was  
performed by the addition of 0.1  $\mu$ M of methotrexate (MTX)  
into the culture media. Amplification was carried out in  
a stepwise fashion with increasing concentration of MTX

(from 0.1 to 3  $\mu$ M). hWI2 purified from amplified clones exhibited identical immunoreactivity as hWI2 obtained from previous non-amplifiable clones.



## WHAT IS CLAIMED IS:

1. A chimeric anti-idiotypic antibody or fragment thereof which specifically binds to the idiotype region of an anti-CEA monoclonal antibody, comprising the rWI2 light chain and heavy chain variable regions, or silent mutations thereof.

2. A humanized anti-idiotypic antibody or fragment thereof which specifically binds the idiotype region of an anti-CEA monoclonal antibody, comprising rWI2 CDR regions and humanized FR regions.

3. An isolated polynucleotide encoding the heavy chain or the heavy chain variable region of a chimeric or humanized antibody or antibody fragment according to claim 1 or 2, comprising sequences encoding at least two rWI2 heavy chain CDRs, selected from the group of CDRs consisting of:

the complementary determining region -1 (CDR-1) sequence NYWMT,

the complementary determining region -2 (CDR-2) sequence SITSTGGTYHAESVKG, and

the complementary determining region -3 (CDR-3) sequence DDYGGQSTYVMDA.

4. An isolated polynucleotide encoding the light chain or the light chain variable region of a chimeric or humanized antibody or antibody fragment according to claim 1 or 2, comprising sequences encoding at least two rWI2 light chain CDRs, selected from the group of CDRs consisting of:

the complementary determining region -1 (CDR1) sequence RASQDIGNYLR,

the complementary determining region -2 (CDR2) sequence GATNLAA, and

the complementary determining region -3 (CDR3) sequence LHHSEYPYT.

5. A chimeric anti-idiotypic antibody according to claim 1, wherein said heavy chain variable region comprises the ratWI2VK sequence shown in Figure 1.

6. A chimeric anti-idiotypic antibody according to claim 1, wherein said light chain variable region comprises the RatWI2VK sequence shown in Figure 2.

7. A humanized anti-idiotypic antibody according to claim 2, wherein the heavy chain variable region comprises the KOLWI2VH-1 or the KOLWI2VH-2 sequence shown in Figure 1.

8. A humanized anti-idiotypic antibody according to claim 2, wherein the light chain variable region comprises the REIWI2VK or the REIWI2VKRS sequence shown in Figure 2.

9. An isolated expression vector comprising a first gene for the WI2 heavy chain and second gene for the WI2 light chain.

10. An isolated expression vector according to claim 9 wherein said light and heavy chains are chimeric or are humanized.

11. A host comprising said expression vector according to claim 9.

12. An isolated first expression vector comprising a gene for WI2 heavy chain and an isolated second expression vector comprising a gene for the WI2 light chain.

13. An isolated first and second expression vectors according to claim 12, wherein said genes are for chimeric or humanized WI2 light and heavy chain.

14. A host comprising said first and second expression vectors according to claim 12.

15. A method of stimulating an immune response in a patient against cancers expressing carcinoembryonic antigen, which comprises administering to said patient an effective amount of a vaccine comprising the humanized anti-idiotypic antibody or antibody fragment of claim 2, conjugated to a soluble immunogenic carrier protein, optionally in combination with a pharmaceutically acceptable vaccine adjuvant.

16. In a method of diagnosis or treatment of a patient, wherein an antibody or antibody fragment that specifically binds CEA is used as a targeting, pre-targeting or therapy agent, either as such or as a component of a conjugate,

the improvement wherein an anti-idiotypic antibody according to claim 2 is used to clear non-targeted antibody or antibody fragment.

17. A method of detecting the presence of an antibody or fragment thereof that specifically binds CEA, in a biological fluid sample, comprising contacting said sample with rWI2, or a chimeric anti-idiotypic antibody or antibody fragment according to claim 1, or a humanized anti-idiotypic antibody or antibody fragment according to claim 2, and detecting binding of said anti-idiotypic antibody or antibody fragment to an antibody idiotype or antibody idiotype fragment in said sample.

18. A method according to claim 16, wherein said anti-idiotypic antibody or antibody fragment is labeled with a radiolabel, an enzyme, or a fluorescent agent.

19. A vaccine, comprising the humanized anti-idiotypic antibody or antibody fragment of claim 2, conjugated to a soluble immunogenic carrier protein, for

use in stimulating an immune response in a patient against a cancer characterized by expression of CEA.

KOL:	V	---	GV	---	R	---	SS	---	I	---	XXXX	V	---	K	---	XXXXXXXXXXXXXX		
RATWI2VH:	QVQL	Q	ESG	DL	VQ	PGR	SLK	LS	CV	AS	GF	T	FS	NY	WT	SIT	STGGGTYHAE	
KOLWI2VH-1:	---	---	GV	---	R	---	SS	---	---	---	---	---	---	---	---	---	SIT	STGGGTYHAE
KOLWI2VH-2:	---	V	---	GV	---	R	---	SS	---	---	---	---	---	---	---	---	SIT	STGGGTYHAE

KOL:	XXXX	---	N	---	F	---	D	---	GV	---	F	---	A	---	XXXXXXXXXXXXXX	---	TP	---		
RATWI2VH:	SVKG	RFT	IS	R	DN	SK	ST	LY	LQ	MN	SL	R	P	E	D	T	AT	Y	WG	QSSVTVSS
KOLWI2VH-1:	SVKG	---	N	---	F	---	D	---	GV	---	---	---	---	---	---	---	---	---	---	---
KOLWI2VH-2:	SVKG	---	N	---	F	---	D	---	GV	---	---	---	---	---	---	---	---	---	---	---

FIG. 1

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REI:  
 RATWI2VK:  
 REIWI2VKRS:  
 REIWI2VK:

---M---	S--S--V	XXXXXXXXXX	Y	--T--A-K	XXXXXX
DIQLTQSPASLPASLGDRVTITC		RASQDIGNYLR	F	QOKPGKSPRLLIY	GATNLAA
---M---	S--S--V	RASQDIGNYLR	-	--T--A-K	GATNLAA
---M---	S--S--V	RASQDIGNYLR	-	--T--A-K	GATNLAA

REI:  
 RATWI2VK:  
 REIWI2VKRS:  
 REIWI2VK:

---	G	--T	-Y	TF--SS-QP--I-T--	XXXXXX	Q	---	QI	T	-T
GVPSRFGS	R	SG	D	SLTINSLESEDMAIYYC	LHHSEYPYT	I	---	GTKLER	K	R
---	---	---	---	TF--SS-QP--I-T--	LHHSEYPYT	-	---	---	---	---
---	G	--T	-	TF--SS-QP--I-T--	LHHSEYPYT	-	---	---	---	---

FIG. 2

oligo 21 caggtccaactgcaggagtcagggggaggtgtagtgacagcctggaa  
[PstI]  
CAGGTCCAACATGCAGGAGTCAGGGGGAGGTGTAGTGCAGCCTGGAAGGTCTCTGAGACTT  
1 -----+-----+-----+-----+-----+-----+-----+ 60  
GTCCAGGTTGACGTCTCTCAGTCCCCCTCCACATCACGTCGGACCTTCCAGAGACTCTGAA [oligo  
K]  
  
Q V Q L Q E S G G G V V Q P G R S L R L -  
|  
(Q to V mutation needed)  
TCCTGTAGCTCATCTCGATTACATTACAGTAATTACTGGATGACTTGGATACGCCAGGCT  
61 -----+-----+-----+-----+-----+-----+ 120  
[oligo K] AGGACATCGAGTAGACCTAAGTGTAAATGACCTACTGAACCTATGCGGTCCGA  
  
S C S S S G F T F S N Y W M T W I R Q A -  
  
[KpnI]  
CCAGGGAAGGGTCTTGAATGGGTTGCGTCCATTACTAGTACTGGTGGTACCTACCAT  
121 -----+-----+-----+-----+-----+ 180  
[oligo K] GGTCCCTTCCAGAACTTACCCACGCGAGTAATGATCATGACCACCACCATGGATGGTA  
ccaacgcaggtaatgatcatgaccaccaccatggatggta  
P G K G L E W V A S I T S T G G G T Y H -  
oligo  
23 atgcagagtcctgtgaaggccgattcactatctccagagataattcaa  
[AlwNI]  
GCAGAGTCTGTGAAGGGCCGATTCACTATCTCCAGAGATAATTCAAAAAACACCCTGTTC  
181 -----+-----+-----+-----+-----+ 240  
CGTCTCAGACACTTCCCGGCTAAGTGATAGAGGTCTCTATTAAGTTTTTGTGGGACAAG [oligo  
L]  
  
cgtctcagacactt (oligo 22)  
A E S V K G R F T I S R D N S K N T L F -  
  
CTGCAATGGACAGTCTGAGGCCTGAGGACACGGGCGTTTATTACTGTTCAAGAGATGAC  
241 -----+-----+-----+-----+-----+ 300  
[oligo L] GACGTTTACCTGTCTCAGACTCCGGACTCCTGTGCCGCAAATAATGACAAGTTCTCTACTG  
  
L Q M D S L R P E D T G V Y Y C S R D D -  
  
[BstEII]  
TACGGAGGACAGAGCACCTATGTTATGGATGCCCTGGGGTCAGGGAACCTCCGGTCACCGTC  
301 -----+-----+-----+-----+-----+ 360  
[oligo L] ATGCCTCCTGTCTCGTGGATAACAATACCTACGGACCCCACTCCCTTGAGGCCAGTGGCAG  
caatacctacggaccccgagcccttgaggccagtgaggag  
Y G G Q S T Y V M D A W G Q G T P V T V -  
  
TCCTCC  
361 ----- 366  
AGGAGG  
aggagg (oligo 24)  
S S -

FIG. 3

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oligo 25 (PvuII)  
gacattgaagctgacccagtcctccatcttccctgtctgcgtctgtgggaga  
oligo 25 atgaccagtcctccatcttccctgtctgcgtctgtgggaga  
GACATTCAGATGACCCAGTCTCCATCTTCCCTGTCTGCGTCTGTGGGAGACAGAGTCACT  
1 -----+-----+-----+-----+-----+ 60  
CTGTAAGTCTACTGGGTCAGAGGTAGAAGGGACAGACCGCAGACACCCTCTGTCTCAGTGA [oligo  
M)  
D I Q M T Q S P S S L S A S V G D R V T -  
ATTACTTGCCGGGCAAGTCAAGACATTGGAATTATTTAAGATGGTTCAGCAGACACCG  
61 -----+-----+-----+-----+-----+ 120  
[oligo M] TAATGAACGGCCCGTTCAGTCTCTGTAACCTTTAATAAATTCTACCAAGGTCGTCTGTGGC  
I T C R A S Q D I G N Y L R W F Q Q T P -  
oligo 27 tggctgcaggggtcccatca  
[PstI]  
GGGAAAGCTCCGAAACTTTTGATTATGGTGCAACCAACTTGGCTGCAGGGGTCCCATCA  
121 -----+-----+-----+-----+-----+ 180  
[oligo M] CCCTTTTCGAGGCTTTGAAAATAAATACCAAGTGGTTGAACCGACGTCCCCAGGGTAGT  
actaaataccacgttggttgaaccgacgtccccagg (oligo 26)  
G K A P K L L I Y G A T N L A A G V P S -  
cggttcagtggcagtggtctggg  
CGGTTTCAGTGGCAGTGGGTCTGGGACAGATTTTACTTTTACCATCTCAAGCCTTCAGCCT  
181 -----+-----+-----+-----+-----+ 240  
[oligo N] GCCAAGTCACCGTCACCCAGACCCTGTCTAAAATGAAAATGGTAGAGTTTCGGAAGTCGGA  
R F S G S G S G T D F T F T I S S L Q P -  
GAAGATATTGCTACTTATTACTGTCTGCACCATCTGAGTATCCATACACGTTTGGAATT  
241 -----+-----+-----+-----+-----+ 300  
[oligo N] CTTCTATAACGATGAATAATGACAGACGTGGTAAGACTCATAGGTATGTGCAAACCTTAA  
tgtgcaaaccttaa  
E D I A T Y Y C L H H S E Y P Y T F G I -  
GGGACCAAGTTGCAGATCAAACGTG  
301 -----+-----+-----+-----+ 325  
[oligo N] CCCTGGTTCAACGTCTAGTTGCAC  
ccctggttcaacgtctagattgcac (oligo 28)  
G T K L Q I K R -

FIG. 4



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# hWI2 BLOCKING ASSAY

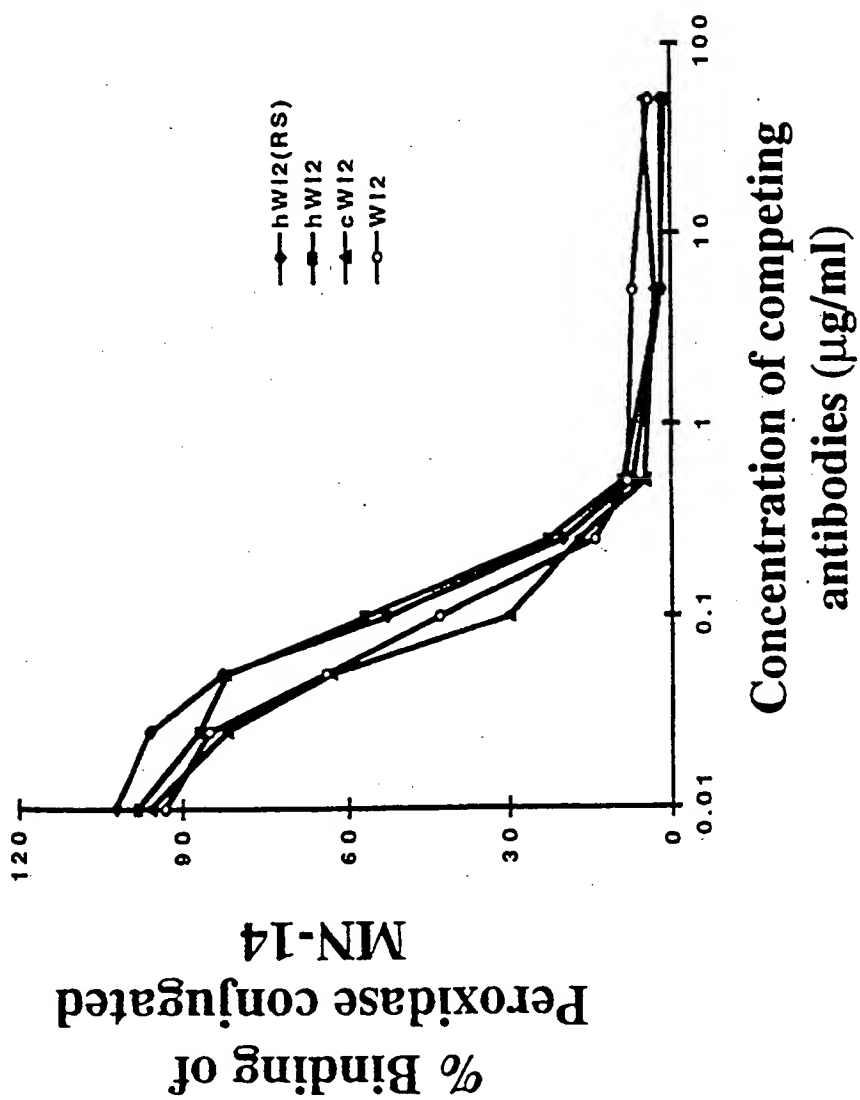


FIG. 5

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GACATTCAGC TGACCCAGTC TCCAGCTTCC CTGCCTGCGT CTCTGGGAGA 50  
CTGTAAGTCG ACTGGGTCAG AGGTCGAAGG GACGGACGCA GAGACCCTCT  
D I Q L T Q S P A S L P A S L G D

CAGAGTCACT ATTACTTGCC GGGCAAGTCA AGACATTGGA AATTATTTAA 100  
GTCTCAGTGA TAATGAACGG CCCGTTTCAGT TCTGTAAACCT TTAATAAATT  
R V T I T C R A S Q D I G N Y L R  
CDR1

GATGGTTCCA GCAGAAACCG GGGAAATCTC CGAGGCTTTT GATTTATGGT 150  
CTACCAAGGT CGTCTTTGGC CCCTTTAGAG GCTCCGAAAA CTAAATACCA  
W F Q Q K P G K S P R L L I Y G

GCAACCAACT TGGCAGCTGG GGTCCCATCA CGGITCAGTC GCAGTAGGTC 200  
CGTTGGTTGA ACCGTCGACC CCAGGGTAGT GCCAAGTCAC CGTCATCCAG  
A T N L A A G V P S R F S G S R S  
CDR2

TGGGTCAGAT TTTTCTCTGA CCATCAACAG CCTGGAGTCT GAAGATATGG 250  
ACCCAGTCTA AAAAGAGACT GGTAGTTGTC GGACCTCAGA CTTCTATACC  
G S D F S L T I N S L E S E D M A

CTATTTATTA CTGTCTGCAC CATTCTGAGT ATCCATACAC GTTTGGAATT 300  
GATAAATAAT GACAGACGTG GTAAGACTCA TAGGTATGTC CAAACCTAA  
I Y Y C L H H S E Y P Y T F G I  
CDR3

GGGACCAAGC TGGAACGGAA ACGG 324  
CCCTGGTTTCG ACCTTGCCTT TGCC  
G T K L E R K R

FIGURE 6

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CAGGTCCAAC	TGCAGGAGTC	AGGGGGAGAT	CTAGTGCAGC	CTGGAAGGTC	50
GTCCAGGTTG	ACGTCCTCAG	TCCCCCTCTA	GATCACGTCG	GACCTTCCAG	
Q V Q L	Q E S	G G D	L V Q P	G R S	
TCTGAAACTT	TCCTGTGTAG	CCTCTGGATT	CACATTCAGT	AATTACTGGA	100
AGACTTTGAA	AGGACACATC	GGAGACCTAA	GTGTAAGTCA	TTAATGACCT	
L K L	S C V A	S G F	T F S	<u>N Y W M</u>	
				CDR1	
TGACTTGGAT	CCGCCAGGCT	CCAGGGGAGG	GTCTTGAATG	GGTTGCGTCC	150
ACTGAACCTA	GGCGGTCCGA	GGTCCCCTCC	CAGAACTTAC	CCAACGCAGG	
<u>T</u> W I	R Q A	P G E G	L E W	V A <u>S</u>	
ATTACTAGTA	CTGGTGGTGG	GACTTACCAT	GCAGAGTCTG	TGAAGGGCCG	200
TAATGATCAT	GACCACCACC	CTGAATGGTA	CGTCTCAGAC	ACTTCCCGGC	
<u>I T S T</u>	<u>G G G</u>	<u>T Y H</u>	<u>A E S V</u>	<u>K G R</u>	
		CDR2			
ATTCACTATC	TCCAGAGATA	ATTCAAAAAG	CACCCTGTAC	CTGCAAATGA	250
TAAGTGATAG	AGGTCTCTAT	TAAGTTTTC	GTGGGACATG	GACGTTTACT	
F T I	S R D N	S K S	T L Y	L Q M N	
ACAGTCTGAG	GCCTGAGGAC	ACGGCCACTT	ATTACTGTTC	AAGAGATAGAC	300
TGTCAGACTC	OGGACTCCTG	TGCCGGTGAA	TAATGACAAG	TTCTCTACTG	
S L R	P E D	T A T Y	Y C S	R <u>D D</u>	
TACGGAGGAC	AGAGCACCTA	TGTTATGGAT	GCCTGGGGTC	AGGGATCTTC	350
ATGCCTCCTG	TCTOGTGGAT	ACAATACCTA	CGGACCCAG	TCCCTAGAAG	
<u>Y G G O</u>	<u>S T Y</u>	<u>V M D</u>	<u>A W G Q</u>	<u>G S S</u>	
		CDR3			
GGTCACCGTC	TCCTCA				376
CCAGTGGCAG	AGGGGT				
V T V	S S				

FIGURE 7

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US97/04696

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : 424/159.1; 530/388.3, 395; 435/252.33, 7.9; 536/23.53

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/159.1; 530/388.3, 395; 435/252.33, 7.9; 536/23.53

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	IRVINE et al. Induction of Delayed-Type Hypersensitivity Responses by Monoclonal Antiidiotypic Antibodies to Tumor Cells Expressing Carcinoembryonic Antigen and Tumor Associated Glycoprotein-72. Cancer Immunology Immunotherapy. May 1993, Vol. 36, No. 5, pages 281-292, whole article.	15-16, 19
Y	PIMM et al. Determination of the Immunoreactivity of Radiolabelled Monoclonal Antibody Fragments by binding to Immobilized Antiidiotypic Antibodies. Life Sciences. 1995. Vol. 56, No. 17, pages 1401-1406, whole article.	16-18

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"A" document member of the same patent family
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"P" document published prior to the international filing date but later than the priority date claimed	

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15 MAY 1997

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US97/04696

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	GAIDA et al. A Monoclonal Antiidiotypic Antibody Bearing the Image of an Epitope Specific to the Carcinoembryonic Antigen. International Journal of Cancer. 28 May 1992. Vol 51, No. 3, pages 459-465, whole article.	1
Y	Morrison. Genetically Engineered (Chimeric) Antibodies. Hospital Practice. 15 October 1989, pages 65-69, 72-74, 77-78, 80, see entire document.	1

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US97/04696

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☒ Claims Nos.: 2-14  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:  
  
The claims recited sequences for which a CRF or a paper copy was not submitted.
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☒ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐

The additional search fees were accompanied by the applicant's protest.

☐

No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US97/04696

## A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

A61K 39/42; C12N 1/20; C12P 21/08; C07H 21/04; C07K 1/00, 16/00; G01N 33/53, 33/542

## B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

Medline, Biosis, Cancerlit, Scisearch

search terms: antiidiotypic antibodies, Carcinoembrionic antigen, cancer treatment or therapy, diagnosis using antiidiotypic antibodies for CEA

## BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

Group I, claim(s) 1-2, 5-8, 15-16, 18-19, drawn to an antiidiotypic antibody and vaccine composition thereof.

Group II, claim(s) 3-4, 9-14, drawn to nucleic acid and expression vector thereof.

Group III, claim 17, drawn to a method of detection of an antibody.

The inventions listed as Groups I-III do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons:

The technical feature linking groups I-III appear to be that they all relate to an antiidiotypic antibody, nucleic acid encoding antibodies and methods to use the antiidiotypic antibody in diagnosis and treatment of cancers.

However, antiidiotypic antibodies for diagnosis and therapy are well known in the prior art and can be used to purify idiotypic antibodies that may have relevant functions in treatment of cancer. Therefore, the technical feature of the linking groups I-III does not constitute a special technical feature as defined by PCT Rule 13.2, as it does not define a contribution over prior art.

The special technical feature of Group I is drawn to a protein/antibody product, and a method of using the antibody product in diagnosing and treating cancers.

The special technical feature of Group II claims are drawn to the nucleic acids and expression vector.

The special technical feature of Group III claims is drawn to a detection method.

Accordingly, Groups I-III are not so linked by the same or a corresponding special technical feature as to form a single general inventive concept.